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## TOXICOLOGICAL EVALUATIONS OF GLYPHOSATE IN ZEBRAFISH EARLY-LIFE STAGES

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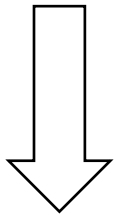
# GLYPHOSATE

*Is a systemic and non-selective post emergence foliar herbicide.  
It is today known as the most widely used herbicide worldwide.*

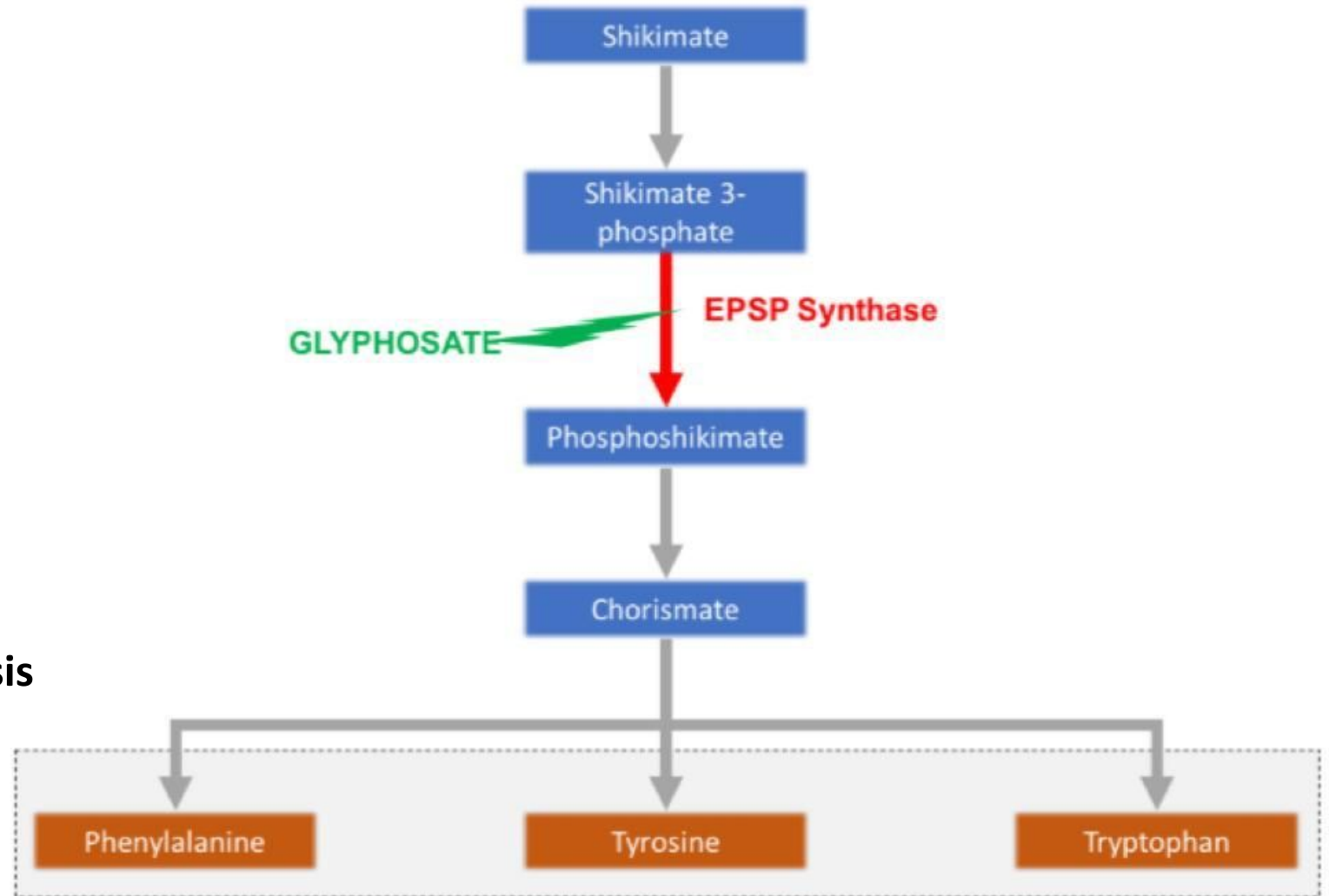


# ***MECHANISM OF ACTION:***

Glyphosate operates via 5-enolpyruvylshikimate-3-phosphate synthase (EPSPS) enzyme, inactivating him.



- Loss of essential amino acids
- Interferes with chlorophyll photosynthesis
- Accumulation of toxic intermediates



# ***THE AIM OF THE STUDY***

Investigate the glyphosate potential effects on the development of zebrafish early-life stages.



## **Embryo**

- Rapid development
- Optical clarity
- Genetic malleability
- External development



## **Larva**

- Optical clarity
- Genetic malleability
- High throughput screening



## **Adult**

- Vertebrate neural structure
- Low cost, easy maintenance
- Genome comprises of several orthologs of human genes mutated in FAD



# ***MATERIALS AND METHODS:***



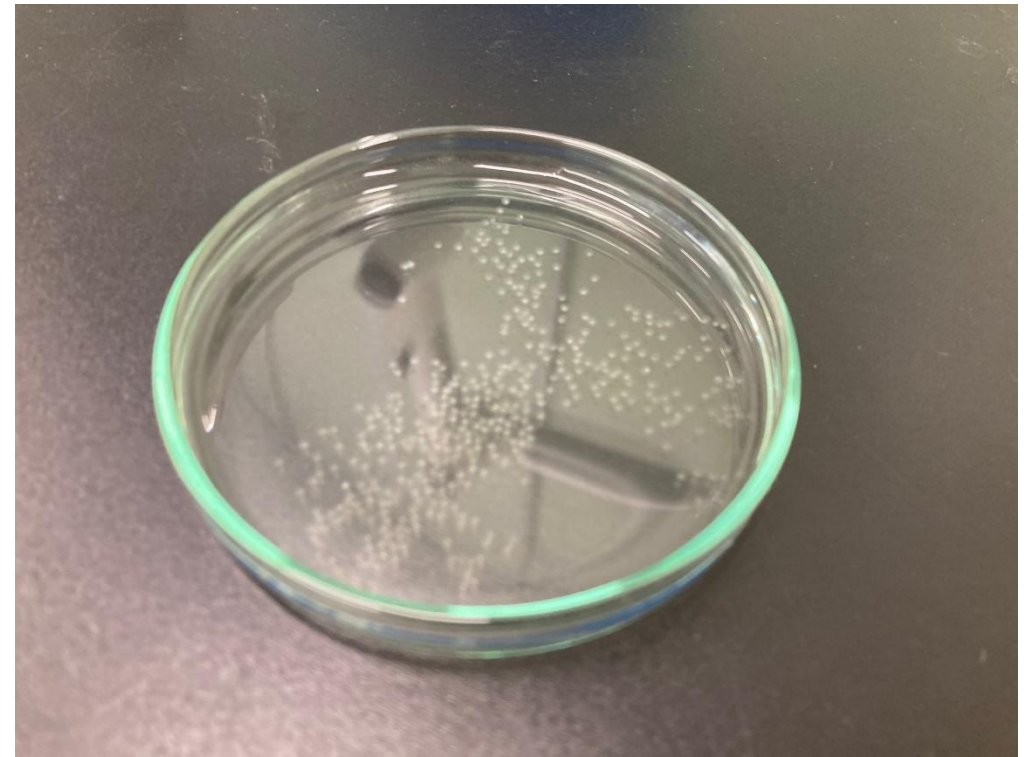
**Zebrafish early-life stages used in the experiments were obtained from the University of Teramo facility. Adult wild type AB strain zebrafish were breed in a recirculating water system with a light/dark cycle**



# ***MATERIALS AND METHODS:***



**Breeding tanks**



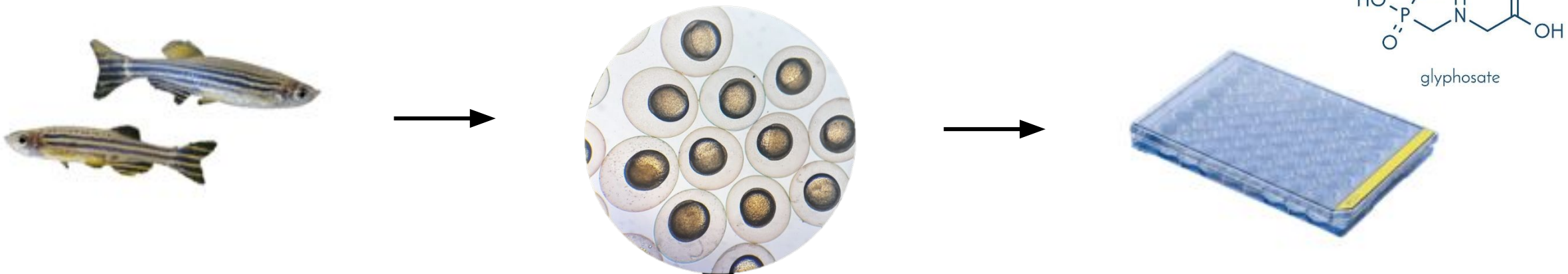
**Collected eggs**



# ***MATERIALS AND METHODS:***

## **Fish Embryo Acute Toxicity Tests (FET)**

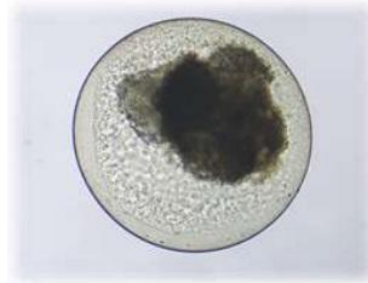
***Tested concentrations: 25, 50, 100, 110, 125 mg/L***  
***Negative control (NK) and positive control***



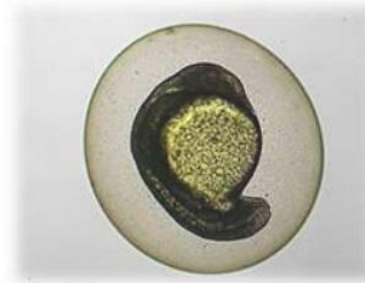
# ***MATERIALS AND METHODS:***



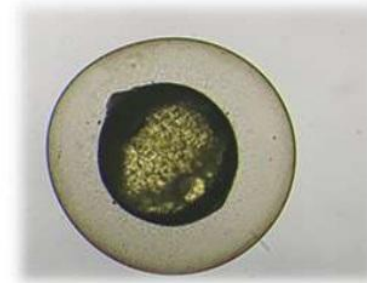
***Zebrafish early-life stages were daily observed up to 96 h with the inverted optical microscope considering four lethal alterations:***



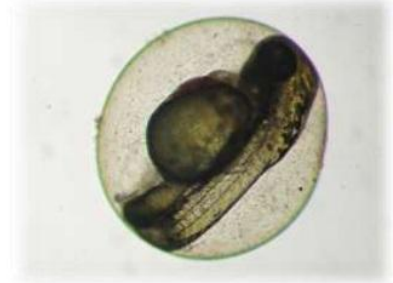
*Coagulation  
of embryos*



*Lack of  
detachment of  
the tail from the  
yolk sac*

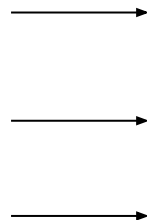


*Lack of the  
somite  
formation*



*Abstance of  
heartbeat*

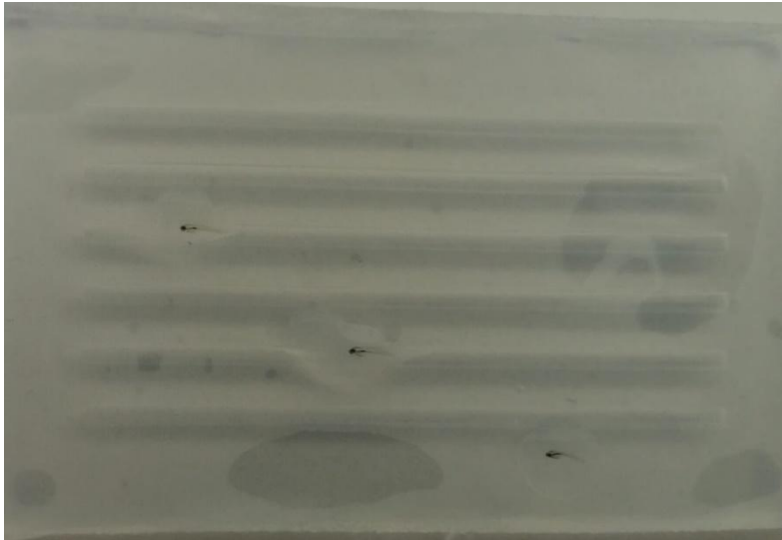
***At the end of the exposure period were evaluated:***





# ***MATERIALS AND METHODS:***

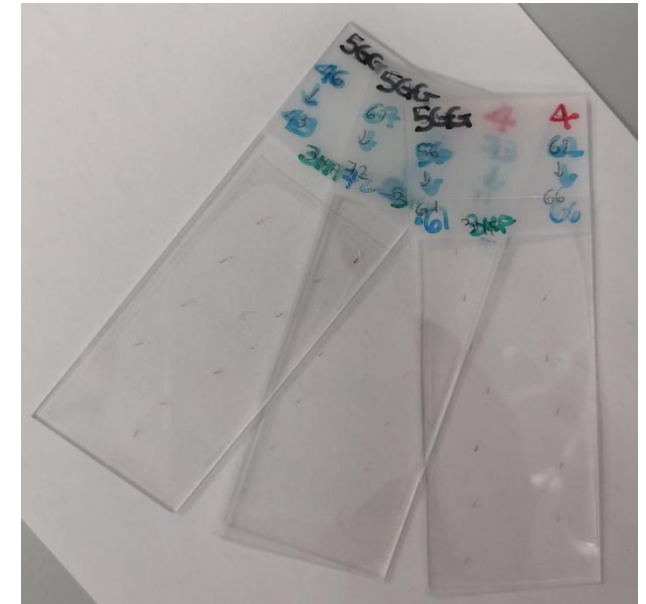
## **Histological analyses**



**Survived larvae at 96 hpf  
treated with 50, 100, 110 and  
125 mg/L**



**Were fixed in 10%  
neutral-buffered formalin  
for 2h**



**5  $\mu$ m thickness sections were  
stained with hematoxylin and  
eosin and then visualized by  
light microscope**

# RESULTS:

## Fish Embryo Acute Toxicity Tests (FET)

| Critical Doses [mcg] |              |   | 0-24 h    | 0-48 h    | 0-72 h    | 0-96 h    |
|----------------------|--------------|---|-----------|-----------|-----------|-----------|
| Survival             | LD10         | ✓ | 88,096    | 86,990    | 86,693    | 85,683    |
|                      | 95%-CL lower |   | n.d.      | n.d.      | n.d.      | n.d.      |
|                      | upper        |   | n.d.      | n.d.      | n.d.      | n.d.      |
| 95%-CL               | LD20         | ✓ | 101,452   | 98,571    | 97,671    | 96,974    |
|                      | lower        |   | n.d.      | n.d.      | n.d.      | n.d.      |
|                      | upper        |   | n.d.      | n.d.      | n.d.      | n.d.      |
| 95%-CL               | LD50         | ✓ | 132,904   | 125,194   | 122,696   | 122,888   |
|                      | lower        |   | n.d.      | n.d.      | n.d.      | n.d.      |
|                      | upper        |   | n.d.      | n.d.      | n.d.      | n.d.      |
| Survival             | LOED         |   | >125,000  | >125,000  | >125,000  | >125,000  |
|                      | NOED         |   | >=125,000 | >=125,000 | >=125,000 | >=125,000 |

# ***RESULTS:***

## **Sublethal alterations:**



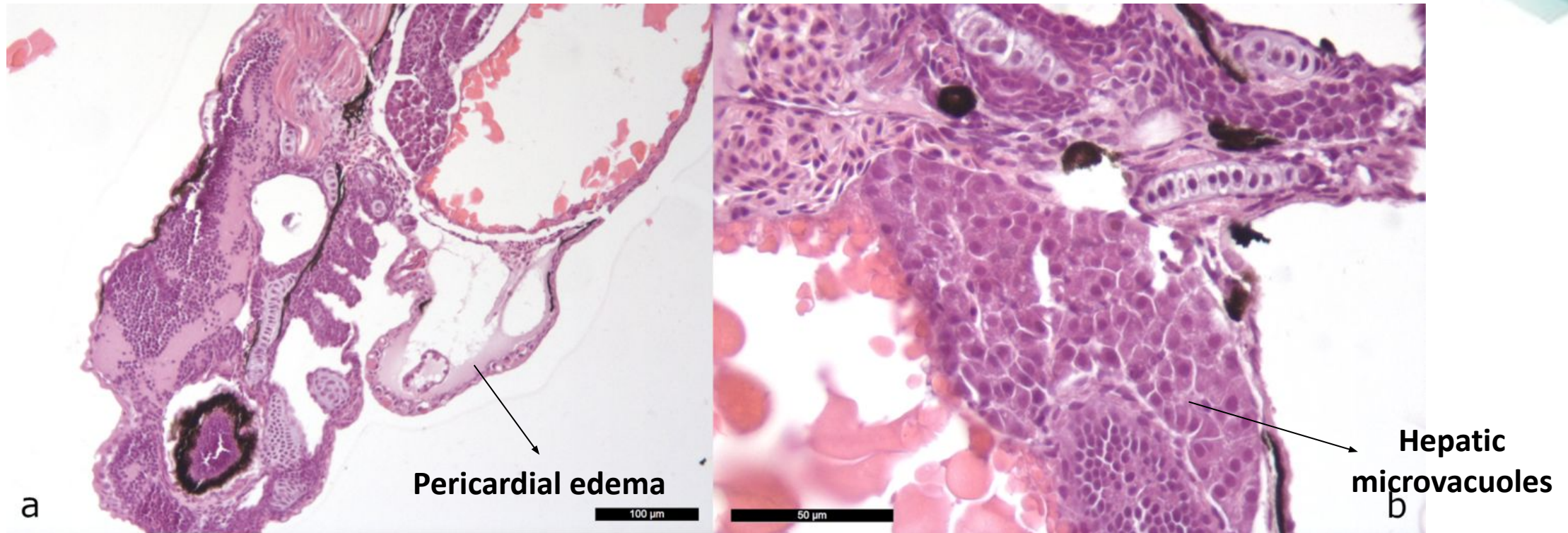
**a-b) From the concentration of 100 mg/L at 48 hpf, the larvae showed pericardial edema and impaired blood flow and blood stasis. In addition, a great number of zebrafish larvae showed a smaller head than NK, suggesting a possible neurological involvement**

**c) Zebrafish larva exposed to negative control (NK)**



# ***RESULTS:***

## **Histological analyses**



**Histological observation confirmed the results of FET test in fact 80% of embryos treated with glyphosate 100 mg/ L showed high pericardial edema and diffuse hepatic microvacuoles. In addition, changes in head size were also observed**

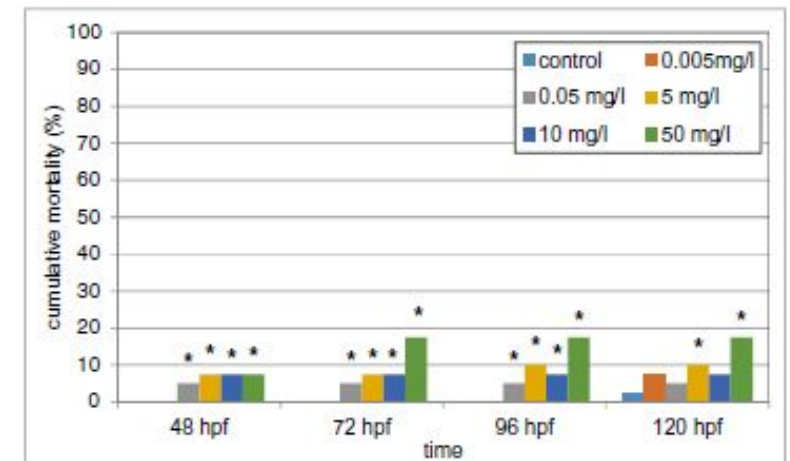
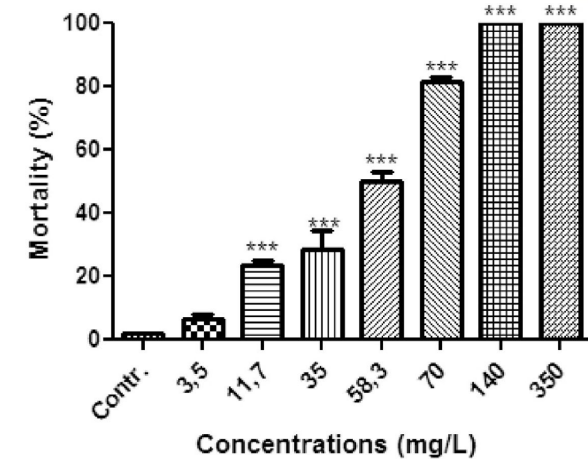
# DISCUSSION:

## TOXICOLOGICAL ENDPOINTS

Panetto, S., Gomes, F., Fraga Gomes, D.S., Campos, Eldo., Romeiro, N.C., Costa, E.P., do Carmo, P.R.L., Feitosa, N.M., & Moraes, J. (2019). **The effects of Roundup® in embryo development and energy metabolism of the zebrafish (*Danio rerio*).** *Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology*, volume 222. doi:0.1016/j.cbpc.2019.04.007.

Rodrigues, L., Gonçalves Costa, G., Lundgren Tha, E., da Silva, L.R., de Oliveira, R., Morais Leme, D., Cestari, M.M., Grisolia, C.K., Campos Valadares, M., & de Oliveira, G.A.R. (2019). **Impact of the glyphosate-based commercial herbicide, its components, and its metabolite AMPA on non-target aquatic organisms.** *Mutation Research/Genetic Toxicology and Environmental Mutagenesis*, volume 842. <https://doi.org/10.1016/j.mrgentox.2019.05.002>

Fiorino, E., Sehonova, P., Plhalova, L., Blahova, J., Svobodova, Z., & Faggio, C. (2018). **Effects of glyphosate on early life stages: comparison between *Cyprinus carpio* and *Danio rerio*.** *Environmental science and pollution research*, 25(9):8542-8549. doi: 10.1007/s11356-017-1141-5



# DISCUSSION:

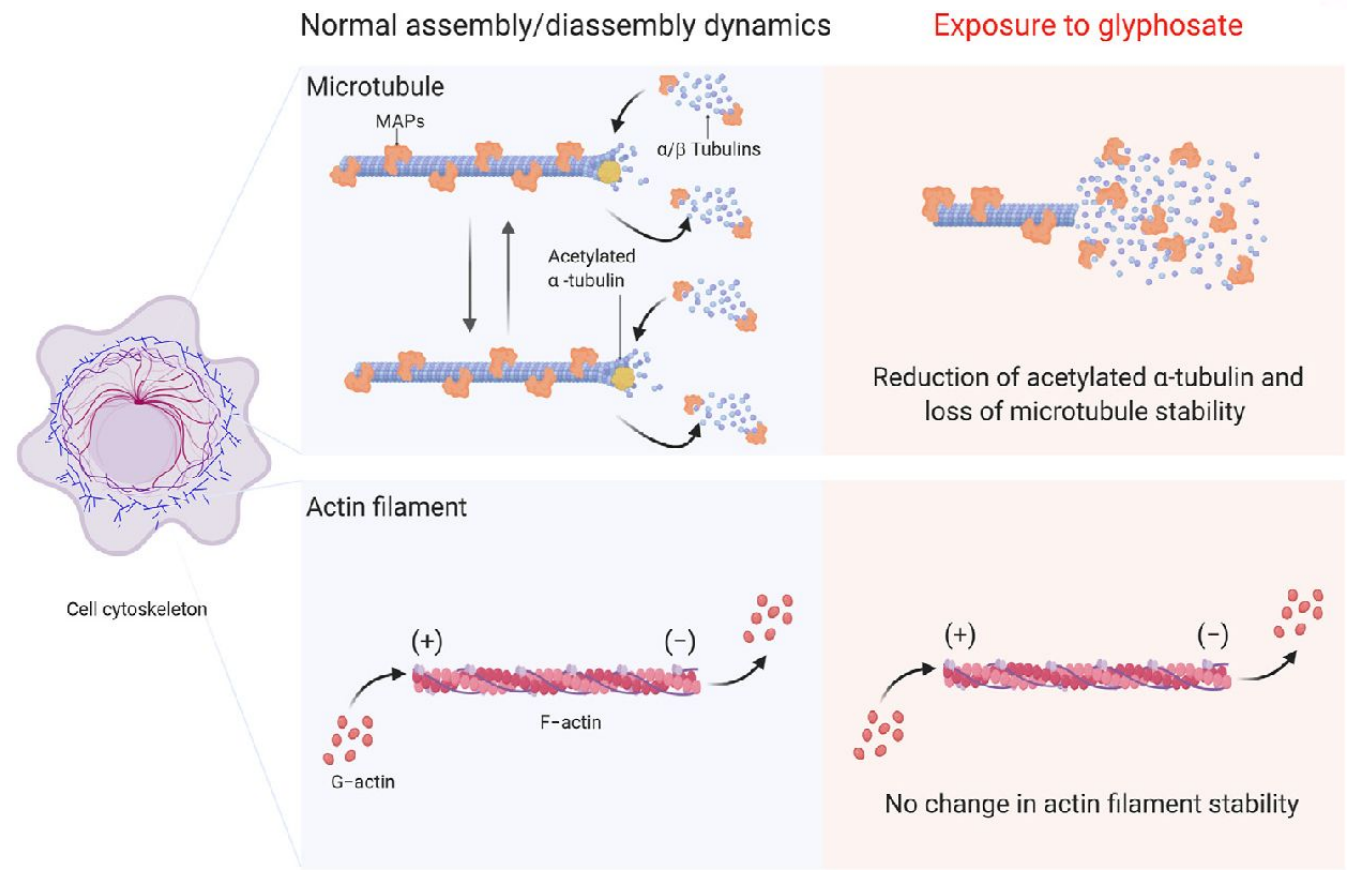
## CARDIOTOXICITY

Lu, J., Wang, W., Zhang, C., Xu, W., Chen, W., Tao, L., Li, Z., Cheng, J., & Zhang, Y. (2022). **Characterization of glyphosate-induced cardiovascular toxicity and apoptosis in zebrafish.** *Science of the Total Environment*, volume 851, Part 2.  
<https://doi.org/10.1016/j.scitotenv.2022.158308>

Diaz-Martin, R.D., Valencia- Hernández, J.D., Betancourt-Lozano, M., & Yáñez-Rivera, B. (2021). **Changes in microtubule stability in zebrafish (*Danio rerio*) embryos after glyphosate exposure.** *Biochemistry, Molecular and Cell Biology*, Volume 7, Issue 1.  
<https://doi.org/10.1016/j.heliyon.2021.e06027>

Sulukan E., Köktürk M., Ceylan H., Beydemir Ş., Işık M., Atamanalp M., & Ceyhun S.B. (2017). **An approach to clarify the effect mechanism of glyphosate on body malformations during embryonic development of zebrafish (*Danio rerio*).** *Chemosphere*, 180:77-85. doi: 10.1016/j.chemosphere.2017.04.018.

## Dysregulation myocyte enhancer factor-2 (MEF2)





# DISCUSSION:

## NEUROTOXICITY

Roy, N. M., Carneiro, B., & Ochs, J. (2016). **Glyphosate induces neurotoxicity in zebrafish.** *Environmental Toxicology and Pharmacology*, 42, 45–54.

<https://doi.org/10.1016/j.etap.2016.01.003>

Diaz-Martin, R.D., Carvajal-Peraza, A., Yáñez-Rivera, B., & Betancourt-Lozano, M. (2021). **Short exposure to glyphosate induces locomotor, craniofacial, and bone disorders in zebrafish (*Danio rerio*) embryos.** *Environmental Toxicology and Pharmacology*, volume 87.

<https://doi.org/10.1016/j.etap.2021.103700>

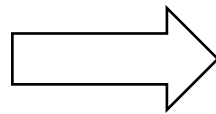
Faria, M., Bedrossiantz, J., Ramirez Rosas, J.R., Mayol, M., Garcia Heredia, G., Bellot, M., Prats, E., Reyero-Garcia, N., Gomez-Canela, C., Gomez-Olivan, L.M., & Raldua D. (2021). **Glyphosate targets fish monoaminergic systems leading to oxidative stress and anxiety.** *Environment International*, volume 146.

<https://doi.org/10.1016/j.envint.2020.106253>

**Reduction of essential genes for the normal development of the central nervous system, such as pax2, pax6, Otx2, ephA4**

# ***CONCLUSION:***

**TOXIC**



Long-terms effects of  
glyphosate

THANK YOU FOR YOUR ATTENTION  
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